



Galactose Oxidase Assay

Method

The reaction velocity is measured in a peroxidase/o-tolidine coupled system as an increase in A425 resulting from the oxidation of galactose. One unit results in a change in A425 of 1.0 per minute at 25° C and pH 6.0 under the defined conditions.

Reagents

- 1) 0.1 M Potassium phosphate buffer, pH 6.0
- 2) 0.5% o-tolidine. Note: o-tolidine has been reported to be carcinogenic. Handle with care.
- 3) Peroxidase. Dissolve at a concentration of approximately 60 u/ml in reagent grade water.
- 4) 10% galactose. Allow to come to equilibrium of mutarotation by allowing to stand overnight.

Enzyme

Dissolve at a concentration of 1 mg/ml in reagent grade water. Dilute further for assay to a concentration of 0.2 - 0.5 units/ml.

Procedure

- 1) Adjust spectrophotometer to 425 nm and 25° C.
- 2) Prepare tolidine-buffer mixture by adding 0.1 ml tolidine to 12 ml 0.1 M potassium phosphate buffer pH 6.0.
- 3) Pipette into each cuvette as follows:

Tolidine-buffer solution	1.7 ml
10% Galactose	1.5 ml
Peroxidase	0.1 ml

Incubate in spectrophotometer at 25° C for 3 - 4 minutes to achieve temperature equilibration and establish blank rate, if any. Add 0.1 ml of appropriately diluted enzyme and record increase in A425/min. from initial linear portion of the curve.



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Calculation

$$\frac{\text{Units}}{\text{mg}} = \frac{\frac{\Delta A_{425}}{\text{min}}}{\text{mg enzyme in reaction mixture}}$$