



3-Phosphoglyceric Phosphokinase

PRINCIPLE:

$\text{GAP} + \beta\text{-NAD} + \text{P}_i \rightarrow (\text{GAPDH}) \rightarrow 1,3\text{-Diphosphoglycerate} + \beta\text{-NADH}$

$1,3\text{-Diphosphoglycerate} + \text{ADP} \rightarrow (3\text{PGK}) \rightarrow 3\text{-Phosphoglycerate} + \text{ATP}$

Abbreviations used:

GAP=Glyceraldehyde 3-Phosphate

$\beta\text{-NAD}$ = $\beta\text{-Nicotinamide Adenine Dinucleotide, Oxidized Form}$

P_i = Inorganic Phosphate

GAPDH=Glyceraldehyde-3-Phosphate Dehydrogenase

$\beta\text{-NADH}$ = $\beta\text{-Nicotinamide Adenine Dinucleotide, Reduced Form}$

ADP = Adenosine 5-Diphosphate

3-PGK=3-Phosphoglyceric Phosphokinase

4-ATP = Adenosine 5'-Triphosphate

CONDITIONS:

T=25°C, pH 6.9, A340nm, Light path=1 cm

METHOD:

Continuous Spectrophotometric Rate Determination

REAGENTS:

A.100 mM Potassium Phosphate Buffer, pH 6.9 at 25°C

(Prepare 100 ml in deionized water using Potassium Phosphate, Monobasic, Anhydrous. Adjust to pH 6.9 at 25°C with 1 M KOH.)

B.25 mM DL-Glyceraldehyde-3-Phosphate Solution (GAP)

(Prepare 1 ml in deionized water using DL-Glyceraldehyde 3-Phosphate, Free Acid.)



C.20 mM B-Nicotinamide Adenine Dinucleotide, Oxidized Form Solution (β - NAD)

(Prepare 1 ml in deionized water using β -Nicotinamide Adenine Dinucleotide.)

D.10 mM Adenosine 5'-Diphosphate Solution (ADP)

(Prepare 1 ml in deionized water using Adenosine 5-Diphosphate, Sodium Salt. Prepare Fresh.)

E. 50 mM Magnesium Sulfate Solution (MgSO₄)

(Prepare 10 ml in deionized water using Magnesium Sulfate, Heptahydrate.)

F. 400 mM Glycine Solution (Glycine)

(Prepare 25 ml in deionized water using Glycine, Free Base.)

G. Glyceraldehyde-3-Phosphate Dehydrogenase Solution (GAPDH)

(Immediately before use, prepare a solution containing 20 units/ml of Glyceraldehyde 3-Phosphate Dehydrogenase in cold deionized water.)

H. 3-Phosphoglyceric Phosphokinase Enzyme Solution (3-PGK)

(Immediately before use, prepare a solution containing 0.3 - 0.6 unit/ml of 3-Phosphoglyceric Phosphokinase in cold Reagent A.)

PROCEDURE:

Pipette (in milliliters) the following reagents into suitable cuvettes:

	Test	Blank
Reagent A (Buffer)	1.40	1.40
Reagent B (GAP)	0.10	0.10
Reagent C(B-NAD)	0.05	0.05
Reagent D(ADP)	0.05	0.05
Reagent E(MgSO)	0.25	0.25
Reagent F(Glycine)	1.00	1.00
Reagent G(GAPDH)	0.05	0.05
Mix by inversion and equilibrate to 25.C. Monitor the As40nm until constant, using a suitablythermostatted spectrophotometer. Then add:		
Reagent H (3-PGK)	0.10	——
Reagent A (Buffer)	——	0.10



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Immediately mix by inversion and record the increase in As40nm for approximately 5 minutes. Obtain the AA340nm/min using the maximum linear rate for both the Test and Blank.

CALCULATIONS:

$$\text{Units/ml enzyme} = \frac{(\Delta A_{340\text{nm}}/\text{min Test} - \Delta A_{340\text{nm}}/\text{min Blank})(3)(\text{df})}{(6.22)(0.1)}$$

3= Total volume (in milliliters) of assay

df = Dilution factor

6.22= Millimolar extinction coefficient of β -NADH at 340 nm

0.1= Volume (in milliliter) of enzyme used

$$\text{Units/mg solid} = \frac{\text{units/ml enzyme}}{\text{mg solid/ml enzyme}}$$

$$\text{Units/mg protein} = \frac{\text{units/ml enzyme}}{\text{mg protein/ml enzyme}}$$

UNIT DEFINITION:

One unit will convert 1.0 umole of 1,3-diphosphoglycerate to 3-phosphoglycerate per minute at pH6.9 at 25°C.

FINAL ASSAY CONCENTRATIONS:

In a 3.00 ml reaction mix, the final concentrations are 50 mM potassium phosphate, 0.83 mM glyceraldehyde-3-phosphate, 0.3mM β -nicotinamide adenine dinucleotide, 0.2 mM adenosine 5'-diphosphate, 4.2 mM magnesium sulfate, 133 mM glycine, 1 unit glyceraldehyde-3-phosphate dehydrogenase, and 0.03 - 0.06 unit 3-phosphoglyceric phosphokinase.